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# Complete Genome Sequence of the Marine Cellulose- and Xylan-Degrading Bacterium *Glaciecola* sp. Strain 4H-3-7+YE5

B. Klippel, A. Lochner, D. Bruce, K. Davenport, C. Detter, L. Goodwin, J. Han, S. Han, L. Hauser, M. Land, N. Mikhailova, M. Nolan, L. Pennacchio, S. Pitluck, R. Tapia, T. Woyke, S. Wiebusch, A. Basner, F. Abe, K. Horikoshi, M. Keller, G. Antranikian

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# **Complete Genome Sequence of the Marine Cellulose- and Xylan-Degrading Bacterium *Glaciecola* sp. Strain 4H-3-7+YE-5**

Barbara Klippel<sup>1</sup>, Adriane Lochner<sup>1,2</sup>, David C. Bruce<sup>3</sup>, Karen Walston Davenport<sup>3</sup>, Chris Detter<sup>3</sup>, Lynne A. Goodwin<sup>4</sup>, James Han<sup>4</sup>, Shunsheng Han<sup>3</sup>, Loren Hauser<sup>2</sup>, Miriam L. Land<sup>2</sup>, Natalia Mikhailova<sup>4</sup>, Matt Nolan<sup>4</sup>, Len Pennacchio<sup>4</sup>, Sam Pitluck<sup>4</sup>, Roxanne Tapia<sup>3</sup>, Tanja Woyke<sup>4</sup>, Sigrid Wiebusch<sup>1</sup>, Alexander Basner<sup>1</sup>, Fumiyoshi Abe<sup>5</sup>, Koki Horikoshi<sup>5</sup>, Martin Keller<sup>2</sup>, Garabed Antranikian<sup>1\*</sup>

<sup>1</sup>Institute of Technical Microbiology, Hamburg University of Technology, Hamburg, Germany

<sup>2</sup>Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, USA

<sup>3</sup>Los Alamos National Laboratory, Los Alamos, New Mexico 87545, USA

<sup>4</sup>DOE Joint Genome Institute, Walnut Creek, California 94598, USA

<sup>5</sup>Extremobiosphere Research Center (XBR), Japan Agency for Marine-Earth Science and Technology (JAMSTEC), Yokosuka, Japan

\*Corresponding author. Institute of Technical Microbiology, Hamburg University of Technology, Kasernenstr. 12, D-21073 Hamburg, Germany. Phone: +49 40 42878 3117. Fax: +49 40 42878 2582. Email: [antranikian@tuhh.de](mailto:antranikian@tuhh.de)

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## Genome Announcement

### Complete genome sequence of the marine, cellulose and xylan degrading bacterium *Glaciecola* sp. 4H-3-7+YE-5

Barbara Klippel<sup>1</sup>, Adriane Lochner<sup>1,2</sup>, David C. Bruner<sup>3</sup>, Karen Walston Davenport<sup>3</sup>, Chris Detter<sup>3</sup>, Lynne A. Goodwin<sup>4</sup>, James Han<sup>4</sup>, Shunsheng Han<sup>3</sup>, Miriam L. Land <sup>2</sup>, Natalia Mikhailova<sup>4</sup>, Matt Nolan<sup>4</sup>, Len Pennacchio<sup>4</sup>, Sam Pitluck<sup>4</sup>, Roxanne Tapia<sup>3</sup>, Tanja Woyke<sup>4</sup>, Martin Keller<sup>2</sup>, Sigrid Wiebusch<sup>1</sup>, Alexander Basner<sup>1</sup>, Fumiyoshi Abe<sup>5</sup>, Koki Horikoshi<sup>5</sup>, Garabed Antranikian<sup>1\*</sup>

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## Abstract

*Glaciecola* sp. 4H-3-7+YE-5 was isolated from deep sea sediments at Suruga Bay in Japan and is capable to efficiently hydrolyze cellulose and xylan. The complete genome sequence of *Glaciecola* sp. 4H-3-7+YE-5 revealed several genes encoding putatively novel glycoside hydrolases involved in plant biomass degradation.

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In order to gain insight into the complete gene repertoire of *Glaciecola* sp. 4H-3-7+YE-5, the genome was sequenced at the DOE Joint genome Institute (JGI) using a combination of Illumina (2) and 454 technologies (10). To this end, we constructed and sequenced an Illumina GAii shotgun library which generated 50,060,436 reads totaling 3,804 Mb, a 454 Titanium standard library which generated 233,681 reads and three paired end 454 libraries with an average insert sizes of 10.0 kb, 5.4 kb, and 5.9 kb which generated 272,557 reads totaling 164.4 Mb of 454 data. All general aspects of library construction and sequencing can be found at <http://www.jgi.doe.gov/>. The initial draft assembly contained 55 contigs in 2 scaffolds. The 454 Titanium standard data and the 454 paired end data were assembled together with Newbler, version 2.3 while the Illumina sequencing data was assembled with VELVET, version 0.7.63 (16). Newbler and Illumina VELVET consensus data as well as read pairs in the 454 paired end library were integrated using parallel phrap, version SPS - 4.24 (High Performance Software, LLC). The software Consed (5-7) was used in the following finishing process. Illumina data was used to increase consensus quality using the software Polisher (Alla Lapidus, unpublished). Mis-assemblies were corrected using gapResolution (Cliff Han, unpublished), Dupfinisher (9), or sequencing cloned bridging PCR fragments. Gaps between contigs were closed by editing in Consed, by PCR and by Bubble PCR (J-F Cheng, unpublished) primer walks. A total of 209 additional reactions were necessary to close gaps and to raise the quality of the finished sequence. The total size of the genome is 5,393,591 bp and the final assembly is based on 137.8 Mb of 454 draft

data which provides 25.6x average genome coverage and 1,774 Mb of Illumina draft data which provides 329x average genome coverage.

The genome of *Glaciecola* sp. 4H-3-7+YE-5 is contained within one large chromosome (5,052,309 bp) and one plasmid (pGLAAG01, 341,282 bp). The complete genome has a total G+C content of 44% and comprises 4,548 predicted protein-encoding genes.

This is the first complete genome sequence for a member of the *Glaciecola* genus and analysis revealed the presence of numerous genes encoding carbohydrate active enzymes like glycoside hydrolases, glycosyl transferases and carbohydrate esterases, making the organism a promising source for biocatalysts.

**Nucleotide sequence accession number.** The complete chromosome and plasmid sequences of *Glaciecola* sp. 4H-3-7+YE-5 have been deposited in GenBank under accession numbers CP002526 and CP002527.

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## **References**

**LLNL-JRNL-501352**  
**LBNL-5334E**

1. Baik, K. S., Y. D. Park, C. N. Seong, E. M. Kim, K. S. Bae, and J. Chun. 2006. *Glaciecola nitratreducens* sp. nov., isolated from seawater. *Int J Syst Evol Microbiol* **56**:2185-2188.
2. Bennett, S. 2004. Solexa Ltd. *Pharmacogenomics* **5**:433-438.
3. Bowman, J., S. McCammon, J. Brown, and T. McMeekin. 1998. *Glaciecola punicea* gen. nov., sp. nov. and *Glaciecola pallidula* gen. nov., sp. nov.: psychrophilic bacteria from Antarctic sea-ice habitats. *Int J Syst Bacteriol* **48**:1213-1222.
4. Chen, L. P., H. Y. Xu, S. Z. Fu, H. X. Fan, Y. H. Liu, S. J. Liu, and Z. P. Liu. 2009. *Glaciecola lipolytica* sp. nov., isolated from seawater near Tianjin city, China. *Int J Syst Evol Microbiol* **59**:73-76.
5. Ewing, B., L. Hillier, M. C. Wendl, and P. Green. 1998. Base-calling of automated sequencer traces using phred. I. Accuracy assessment. *Genome Res* **8**:175-185.
6. Ewing, B., and P. Green. 1998. Base-calling of automated sequencer traces using phred. II. Error probabilities. *Genome Res* **8**:186-194.
7. Gordon, D., C. Abajian, and P. Green. 1998. Consed: a graphical tool for sequence finishing. *Genome Res* **8**:195-202.
8. Guo, B., X. L. Chen, C. Y. Sun, B. C. Zhou, and Y. Z. Zhang. 2009. Gene cloning, expression and characterization of a new cold-active and salt-tolerant endo-beta-1,4-xylanase from marine *Glaciecola mesophila* KMM 241. *Appl Microbiol Biotechnol* **84**:1107-1115.
9. Han, C. S., and P. Chain (2006). Finishing repetitive regions automatically with Dupfinisher. *Proceedings of the 2006 International Conference on Bioinformatics and Computational Biology*. Las Vegas, NV.
10. Margulies, M., M. Egholm, W. E. Altman, S. Attiya, J. S. Bader, L. A. Bemben, J. Berka, M. S. Braverman, Y. J. Chen, Z. Chen, S. B. Dewell, L. Du, J. M. Fierro, X. V. Gomes, B. C. Godwin, W. He, S. Helgesen, C. H. Ho, G. P. Irzyk, S. C. Jando, M. L. Alenquer, T. P. Jarvie, K. B. Jirage, J. B. Kim, J. R. Knight, J. R. Lanza, J. H. Leamon, S. M. Lefkowitz, M. Lei, J. Li, K. L. Lohman, H. Lu, V. B. Makhijani, K. E. McDade, M. P. McKenna, E. W. Myers, E. Nickerson, J. R. Nobile, R. Plant, B. P. Puc, M. T. Ronan, G. T. Roth, G. J. Sarkis, J. F. Simons, J. W. Simpson, M. Srinivasan, K. R. Tartaro, A. Tomasz, K. A. Vogt, G. A. Volkmer, S. H. Wang, Y. Wang, M. P. Weiner, P. Yu, R. F. Begley, and J. M. Rothberg. 2005. Genome sequencing in microfabricated high-density picolitre reactors. *Nature* **437**:376-380.
11. Matsuyama, H., T. Hirabayashi, H. Kasahara, H. Minami, T. Hoshino, and I. Yumoto. 2006. *Glaciecola chathamensis* sp. nov., a novel marine polysaccharide-producing bacterium. *Int J Syst Evol Microbiol* **56**:2883-2886.



12. Prabakaran, S. R., R. Manorama, D. Delille, and S. Shivaji. 2007. Predominance of *Roseobacter*, *Sulfitobacter*, *Glaciecola* and *Psychrobacter* in seawater collected off Ushuaia, Argentina, Sub-Antarctica. FEMS Microbiol Ecol **59**:342-355.
13. Romanenko, L. A., N. V. Zhukova, M. Rohde, A. M. Lysenko, V. V. Mikhailov, and E. Stackebrandt. 2003. *Glaciecola mesophila* sp. nov., a novel marine agar-digesting bacterium. Int J Syst Evol Microbiol **53**:647-651.
14. Van Trappen, S., T. L. Tan, J. Yang, J. Mergaert, and J. Swings. 2004. *Glaciecola polaris* sp. nov., a novel budding and prosthecate bacterium from the Arctic Ocean, and emended description of the genus *Glaciecola*. Int J Syst Evol Microbiol **54**:1765-1771.
15. Yong, J. J., S. J. Park, H. J. Kim, and S. K. Rhee. 2007. *Glaciecola agarilytica* sp. nov., an agar-digesting marine bacterium from the East Sea, Korea. Int J Syst Evol Microbiol **57**:951-953.
16. Zerbino, D. R., and E. Birney. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res **18**:821-829.
17. Zhang, D. C., Y. Yu, B. Chen, H. X. Wang, H. C. Liu, X. Z. Dong, and P. J. Zhou. 2006. *Glaciecola psychrophila* sp. nov., a novel psychrophilic bacterium isolated from the Arctic. Int J Syst Evol Microbiol **56**:2867-2869.
18. Zhang, Y. J., X. Y. Zhang, Z. H. Mi, C. X. Chen, Z. M. Gao, X. L. Chen, Y. Yu, B. Chen, and Y. Z. Zhang. 2010. *Glaciecola arctica* sp. nov., isolated from Arctic marine sediment. Int J Syst Evol Microbiol, in press.

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**LLNL-JRNL-501352**  
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13. Romanenko, L. A., N. V. Zhukova, M. Rohde, A. M. Lysenko, V. V. Mikhailov, and E. Stackebrandt. 2003. *Glaciecola mesophila* sp. nov., a novel marine agar-digesting bacterium. *Int J Syst Evol Microbiol* **53**:647-651.
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